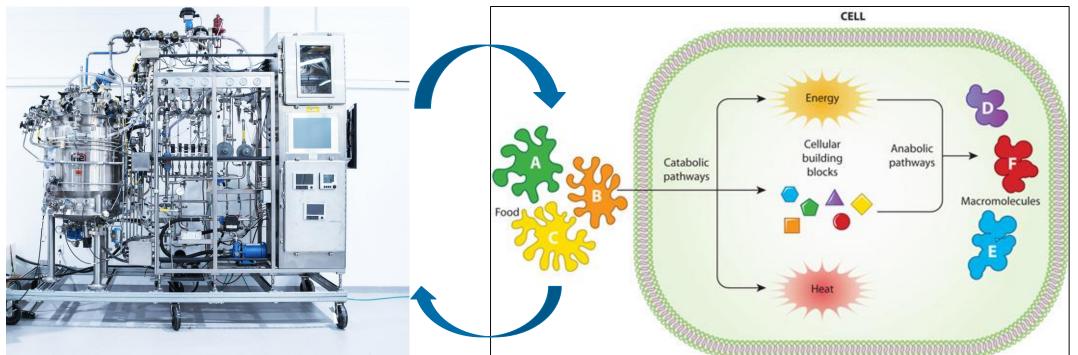
## From fermentation control to protein structure monitoring: the value of Raman spectroscopy as an analysis tool in plant protein ingredients

## INTRODUCTION

Bioreactor parameters affect cellular metabolic processes and final product quality



- Process Analytical Technologies (PAT) provide real-time, in situ process monitoring
- Optimized bioprocess require automated process control
- PAT-based monitoring of bioreactor parameters is the first step toward automated process control

PAT Bioreactor **Traditional Bioreactor** Nutrient concentration determines feed rate Real-time Process Process analytics Understanding Control Extract samples In-line Raman

## **EXPERIMENTAL**

Raman spectroscopy has been an analysis technique for *in situ* fermentation monitoring since 1978 and for understanding biological molecules since 1937! [1-4]



*In situ* bioprocess monitoring achieved using Kaiser probes for lab-scale/PD (bIO-Optic, left) or manufacturing scale (bIO-PRO, right) in single-use, glass or stainless steel bioreactors.

### Rolf Wolthuis<sup>1</sup>, Tomaso Della Vedova<sup>1</sup>, Karen Esmonde-White<sup>2</sup>

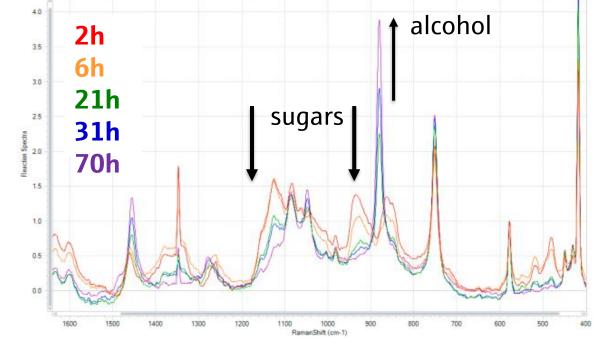
<sup>1</sup> Endress+Hauser | Nikkelstraat 6 | 1411 AJ Naarden | Netherlands <sup>2</sup>Kaiser Optical Systems, Inc. | 371 Parkland Plaza | Ann Arbor, MI 48103 | USA

## **RESULTS AND DISCUSSION**

### Raman spectroscopy: in situ, multi-attribute, real-time bioprocess monitoring

# MeOH "spike" added 49.24 53.2 57.24

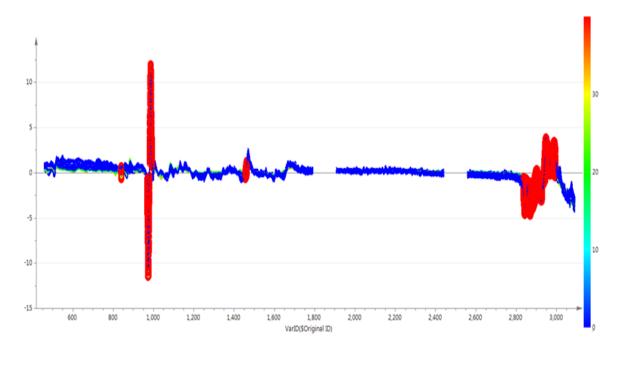
### Fermentation Spectra



### The specificity of *in situ* Raman spectroscopy in bioprocesses enables feedback control and model transfer without significant rework

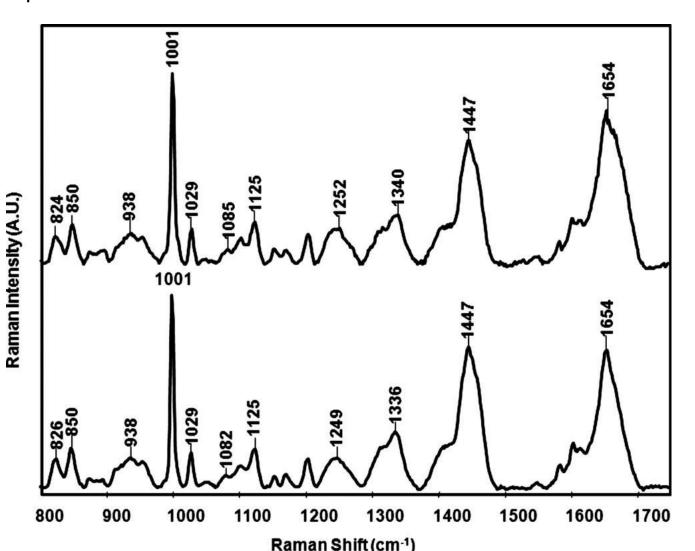
### In situ Raman-based predictions during yeast fermentation

Variable weighing in glycerin spectra for PLS (SIMCA)



### Raman spectroscopy of proteins [4-5]

Raman spectra of proteins is rich with information about the protein backbone and side chains



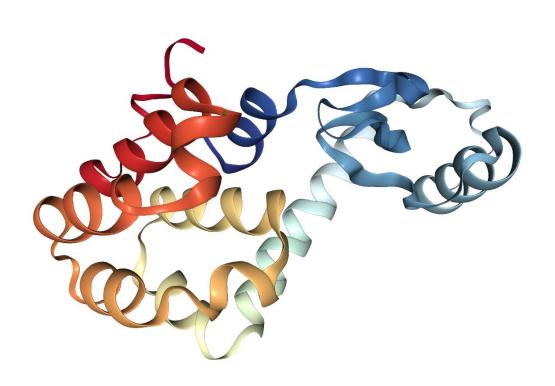
Protein secondary structure such as  $\alpha$ -helix, random coil, and  $\beta$ -sheets are can be understood with a Raman spectrum

Band assignment	Component
Doublet	Tyrosine, Phenylalanine
C-C protein backbone	
Ring breathing	Phenylalanine
C-N, C-C stretch	Protein, polysaccharides
C-C, C-OH C-N stretch C-O-C glycosidic linkage	Protein, polysaccharides
Amide N-H, α-helix	Protein structure
Amide N-H, random coil	Protein structure
$CH_2/CH_3$ wag	Protein
CH <sub>2</sub> /CH <sub>3</sub> deformation	Organic molecules
Amide C=O, α-helix	Protein structure
Amide C=O, random coil	Protein structure
Amide C=O, β-sheet	Protein structure
	Doublet C-C protein backbone Ring breathing C-N, C-C stretch C-C, C-OH C-N stretch C-O-C glycosidic linkage Amide N-H, $\alpha$ -helix Amide N-H, random coil CH <sub>2</sub> /CH <sub>3</sub> wag CH <sub>2</sub> /CH <sub>3</sub> deformation Amide C=O, $\alpha$ -helix Amide C=O, random coil

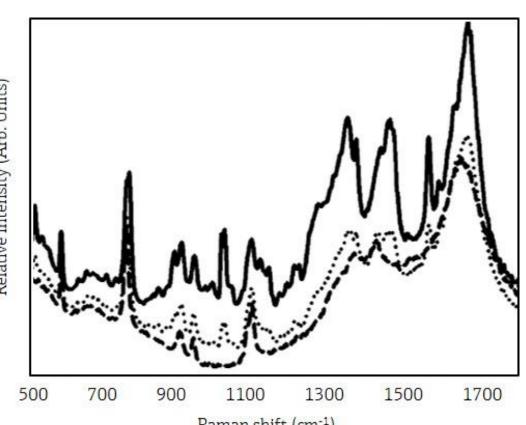
### 5 3 0 15 20 15 20 23 30 VPard20(0)VCESH (g)(1) RM SEE = 2.56006 RMSECV = 3.19131 —— Raman-predicted glycerin Offline measured glycerin man when he when the service when the service of th -10 0 40 60 80 100 120 140 160 120 240 240 240 260 280 800 520 540 860

### Optimization of lysozyme crystallization [6]

Lysozyme was chosen as a model protein to optimize pH, ionic strength, and temperature conditions during crystallization



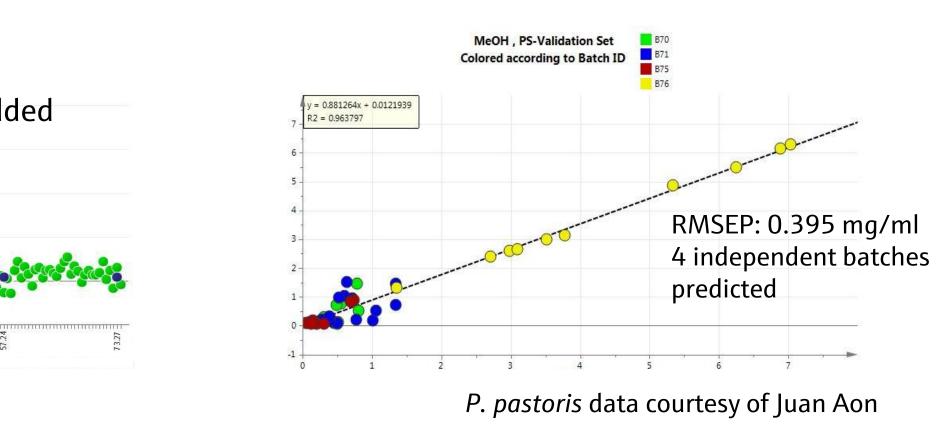
Raman spectra of lysozyme and the acetate buffer solution indicates that meaningful protein information could be obtained in an aqueous environment

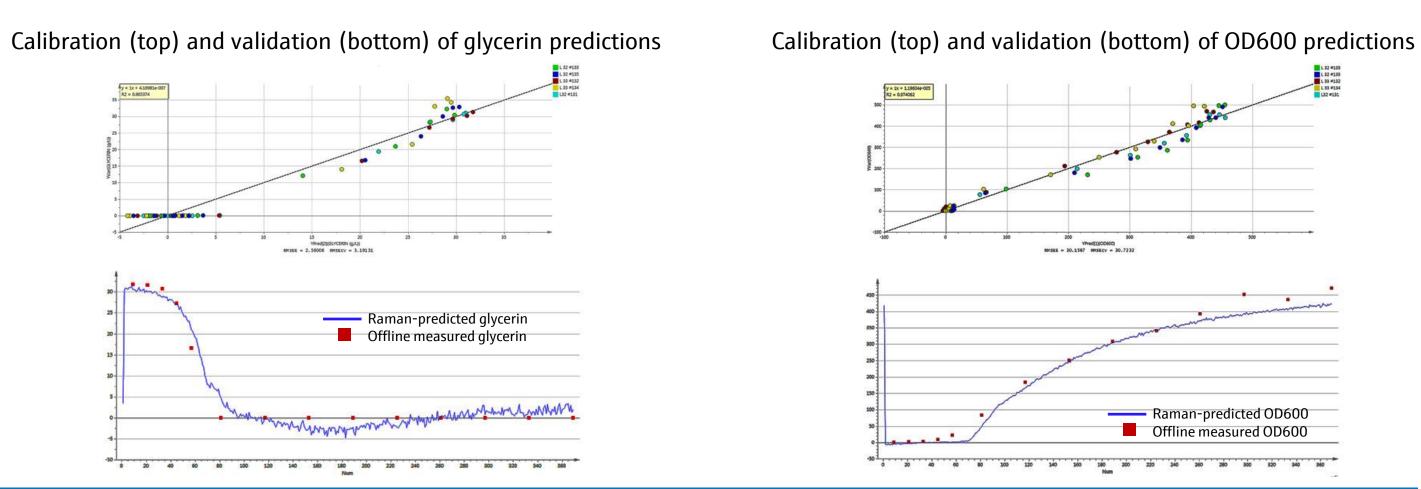


The 155 cm<sup>-1</sup>, 750 cm<sup>-1</sup>, and 2940 cm<sup>-1</sup> bands the crystallization process



### Monitoring and predicting methanol in *Pichia pastoris* fermentation





### Raman: label-free measurements of protein composition and molecular structure

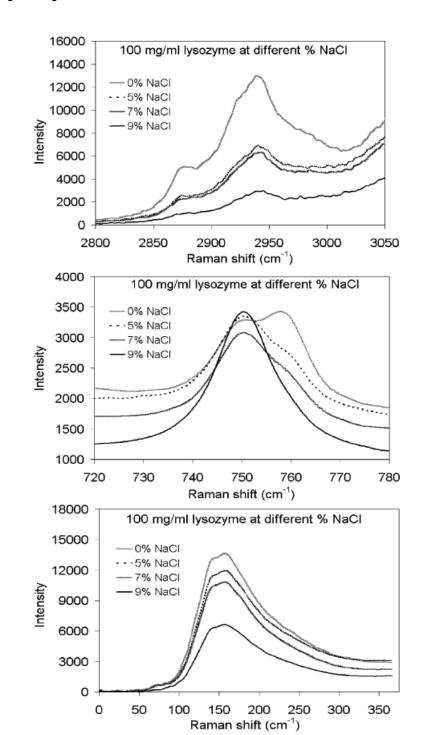
Raman shift (cm<sup>-1</sup>)

were found to be responsive to supersaturation, aggregation, and eventual crystallization during

In the studies described, a Kaiser Raman Rxn1  $(\lambda = 785 \text{ nm})$  was used to collect Raman spectra directly in aqueous lysozyme solutions to monitor crystal growth. (current model: Raman Rxn2, shown below)



### At varying salt concentrations, Raman spectra indicate various states in lysozyme molecular structure







- Post-translational modifications

As a non-destructive, label-free, technique, a Raman-measured protein sample can also be examined by microscopy, mass spectrometry, x-ray crystallography, and rheology. Kaiser Raman is a valuable PAT throughout a product's lifecycle, enabling consistent product quality and process optimization.

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14.	Rowla
15.	Weiss



## CONCLUSIONS

Kaiser Raman is proven as a process analytical technology for industrial bioprocesses. The high chemical specificity of Raman to biological components such as alcohols, sugars, biomass and amino acids enables accurate bioprocess measurements and enables method transfer. *In situ* Raman probes are compatible with autoclave, SIP, CIP and gamma sterilization procedures. Raman spectroscopy has proven benefits for monitoring and control for industrial bioprocesses at the miniature bioreactor, benchtop, pilot, and cGMP manufacturing-scale scales[7-12] to achieve these goals:

- Raman has also been demonstrated in characterizing proteins for these applications **[13-15]** and:
- Crystallization and aggregation
- Higher order structure
- Structure elucidation using isotope exchange
- Molecular response to mechanical stress

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